Conclusive evidence for a distinct congenital stationary night blindness locus in Xp21.1

Arthur A B Bergen, J B ten Brink, F Riemslag, E J M Schuurman, F Meire, N Tijmes, P T V M de Jong

Abstract
X linked congenital stationary night blindness (CSNBX) is a non-progressive retinal disorder characterised by decreased visual acuity and disturbance of night vision. CSNBX appears to be not only clinically but also genetically heterogeneous. On studying a single large family, we recently suggested the presence of a distinct locus for CSNBX in Xp21.1. Here, we describe the results of a linkage analysis in another large CSNBX family, which confirms this finding. Thus, the data presented here provide conclusive evidence for a distinct CSNBX locus in Xp21.1, closely linked to the X linked retinitis pigmentosa type 3 gene. The results combined with other published results indicate the order Xpter-DXS451-DMD-DYS11-(DXS1110, CSNBX1,XLRP3)-DXS7-(CSNBX2, XLRP2)-DXS14-Xcen. (J Med Genet 1996;33:869–872)

Key words: genetic mapping; congenital stationary night blindness; X chromosome; linkage analysis.

Congenital stationary night blindness (CSNB) is a defect in night vision, which is present at birth. Night blindness may occur as a distinct genetic disease entity, or as part of more complex inherited disorders, among them retinitis pigmentosa.

Clinical and genetic heterogeneity has been suggested for both autosomal dominant and X linked CSNB (CSNBX). The clinical heterogeneity of CSNBX in patients, even within the same family, is the result of subtle variation of the involvement of rods or cones, or both, in the disease, which can be visualised through dark adaptation curves and electroretinogram (ERG) amplitudes. Two three clinical forms of CSNBX have been distinguished: complete CSNB (CSNB1), incompletely CSNB (CSNB2), and Aland Island eye disease (AIED). CSNB1 is frequently associated with high myopia. We showed recently that the clinical expression of CSNB1 and CSNB2 appears not to correlate with distinct disease genes (Tijmes et al, manuscript submitted).

So far, three confirmed positions for CSNBX genes have been reported, all proximal to the reference locus DXS7 on proximal Xp. Multiple linkage studies have located a CSNBX gene to the chromosomal interval between DXS7 and DXS426, while another gene has been located even more proximal to the latter chromosomal interval. The localisation of the AIED gene has been refined to between DXS7 and DXS255.

However, we recently suggested that yet another CSNBX gene may be located in Xp21.1, distal to the DXS7 reference locus. Here, we describe a linkage analysis in a new family with CSNBX, thereby providing conclusive evidence for a distinct CSNBX gene locus in Xp21.1.

Subjects and methods
CASES AND CONTROLS
Over the years, all members of pedigree P520 (fig 1) came to our department for ophthalmic diagnosis and genetic counselling. More recently, ophthalmic re-examination was performed, and blood was collected for DNA studies after informed consent. All pedigree members who took part in this study underwent an ophthalmological and electrophysiological examination.

Ophthalmological examination included best refracted visual acuity in both eyes, slit lamp examination, and direct and indirect fundoscopy in full mydriasis. Colour vision was examined with the Panel D-15 (unsaturated) and the Ishihara test. Affected males were examined by kinetic Goldmann perimetry using V-4-e, III-4-e, and I-4-e targets.

Flash ERGs were made, according to the ISCEV standard protocol, with a Ganzfeld dome. Recordings were made in mydriasis after 10 minutes dark adaptation in total darkness. The dark adaptation curves were obtained with a Goldmann Weckers adaptometer. After 10 minutes’ light adaptation (3000 Lux), the threshold to a 15° diameter square wave modulated field, at 11° below the fixation mark, was determined in total darkness.

DNA AND LINKAGE ANALYSIS
Southern and PCR analyses were essentially carried out as previously described. Details
From pedigree P520, five affected males (cases), seven obligate carriers, and three unaffected males (controls) were examined. The cases (aged from 16 to 64 years) had all experienced night blindness from very early childhood onwards. All had a reduced visual acuity which showed no deterioration during lifetime. All cases had myopia (ranging from S-3 to S-9) and a fine horizontal nystagmus.

Fundoscopically, we did not find a pale optic disc, attenuated vessels, bone corpuscles, or retinal pigment epithelium changes in any of the affected males. Visual field tests showed no concentric narrowing or scotomas. Two patients had a slightly concentric restriction of their visual field. However, these patients did not have the field defects seen in retinitis pigmentosa, but a small restriction because of their high myopia (> 9). Colour vision was normal in all members of the pedigree examined.

None of the cases experienced deterioration, during an average follow up period of five years, of their visual acuity or ERG recordings. The ERGs of the patients seemed to lack any rod component, and showed the typical Schubert Bornschein “negative” wave shape, classifying all cases into the category of complete CSNBX. The cone ERG of all cases was not reduced in amplitude, but showed a slightly altered wave shape: the a wave was broader and the b wave was reduced. Dark adaptation showed absence of a rod mediated branch.

The six obligate carriers and one possible carrier had a normal visual acuity, no myopia, and no abnormalities on ERG. The controls also had normal visual acuities and normal ERG. In conclusion, the data from these examinations strongly support the segregation of (complete) CSNBX in this family. Apart from the night blindness, as shown by the dark adaptation curves, there were no clinical or electrophysiological signs of retinitis pigmentosa.

DNA ANALYSIS
The CSNBX family P520 was studied with 11 markers from proximal Xp. Very preliminary data on a few members of this family were reported by Li et al.17

Assuming the fewest number of recombinants, the most likely configuration of alleles is shown in fig 1. Only one locus, DXS1110, cosegregated fully without recombination with the CSNBX gene in this pedigree. A recombination event between CSNBX and the more proximally located loci DXS7/DXS228, DXS1003, DXS426, DXS991, and DXS1125 was shown by the haplotype of patient V.29. A recombination event between CSNBX and the more distally located markers DXS451, DMD49, DMD44, DYS2, and DYS1 could be inferred by comparing the combined haplotypes of IV.15, IV.17, IV.18, and IV.19 with the combined haplotypes of III.4 and III.9, given the assumption that the haplotype of patient III.5, if he had been studied, could possibly have shown a single recombination event. Thus, the most likely gene marker order by haplotype analysis is Xpter-DXS451-DMD-DYS1-DXS1110.
**Table 1** Two point lod scores between CSNBX and a number of key loci in pedigrees P520 and P23340

<table>
<thead>
<tr>
<th>Locus</th>
<th>0.0</th>
<th>0.01</th>
<th>0.05</th>
<th>0.1</th>
<th>0.2</th>
<th>0.3</th>
</tr>
</thead>
<tbody>
<tr>
<td>DSX451</td>
<td>−7.5</td>
<td>−2.14</td>
<td>−1.17</td>
<td>−0.18</td>
<td>0.16</td>
<td></td>
</tr>
<tr>
<td>DMD441</td>
<td>−2.0</td>
<td>−0.16</td>
<td>0.43</td>
<td>0.57</td>
<td>0.54</td>
<td></td>
</tr>
<tr>
<td>DYS1</td>
<td>−3.42</td>
<td>3.72</td>
<td>3.51</td>
<td>2.76</td>
<td>1.86</td>
<td></td>
</tr>
<tr>
<td>DXS1100</td>
<td>3.66</td>
<td>3.58</td>
<td>3.26</td>
<td>2.87</td>
<td>2.07</td>
<td>1.29</td>
</tr>
<tr>
<td>DXS7</td>
<td>−1.26</td>
<td>2.27</td>
<td>2.58</td>
<td>1.98</td>
<td>1.33</td>
<td></td>
</tr>
<tr>
<td>DXS1003</td>
<td>−0.12</td>
<td>1.31</td>
<td>1.73</td>
<td>1.66</td>
<td>1.19</td>
<td></td>
</tr>
<tr>
<td>DXS426</td>
<td>−1.83</td>
<td>−0.01</td>
<td>0.54</td>
<td>0.72</td>
<td>0.52</td>
<td></td>
</tr>
<tr>
<td>DXS14</td>
<td>−3.13</td>
<td>−0.88</td>
<td>0.01</td>
<td>0.19</td>
<td>0.46</td>
<td></td>
</tr>
</tbody>
</table>

CSNBX1-XLRP3)-DXS7-DXS1003-DXS426-DXS991-DXS1125-Xcen.

The results of the two point linkage analyses of pedigree P520 and pedigree P23340, which has previously been described by us, have been presented in table 1.

Multipoint lod scores were carried out with different sets of markers. Two of the most informative results are shown. The position of CSNBX was independently tested against the “key” loci DXS7 (fig 2) and DYS1 (fig 3) on a background map of loci flanking these loci.

The results shown in fig 2 indicate that the CSNBX gene in families P520 and P23340 is localised distal to DXS7, with a probability of more than 1000:1. Since the DYSII locus and CSNBX yielded highly significant two point lod scores (table 1), it was of interest to locate CSNBX relative to this locus as well. Fig 3 shows that CSNBX most likely is localised proximal to DYSII with a probability of at least 100:1. If DXS1110 was included in the analysis, multipoint lod scores favouring the interval DYSII-DXS228 as the most likely chromosomal region for CSNB exceeded 6.1, while other positions of CSNBX became even less likely.

**Discussion**

We previously suggested the presence of a distinct CSNBX gene in band Xp21.1 of the human X chromosome. However, that hypothesis was based on an observation in a single pedigree. A rare chromosomal rearrangement, a double crossover or another genetic mechanism leading to biased results in that pedigree, could not be excluded.

In this paper, we report a second family in which the CSNBX gene maps to Xp21.1. Combining our results with our previously published Xp21.1 linked pedigree, we present here conclusive evidence for a second CSNBX locus on the short arm of the human X chromosome. Our data, combined with other published data, yield the chromosomal gene/marker order Xpter-DXS451-DMD-DYS1-DXS1110-CNBX1-XLRP3)-DXS7-(CSNBX2, XLRP2)-DXS14-Xcen.

Interestingly, the chromosomal positions of the CSNBX loci appear to coincide with those previously found for X linked retinitis pigmentosa. The latter finding provides further evidence for a functional relationship between RP and CSNB, and, in addition, may have important consequences for clinical investigation, gene mapping, and positional cloning studies of CSNBX and XLRP.

A functional relationship between CSNBX and RP was previously suggested by the finding that both CSNB and RP were shown to be caused by different DNA mutations in the rhodopsin gene (chromosome 3q) as well as in the B subunit of phosphodiesterase (chromosome 4p). Our data suggest a positional relationship between CSNBX and XLRP and support that hypothesis.

From a clinical and functional point of view, it would be of interest to investigate whether CSNBX is truly stationary, or, at least in some patients, may be slightly progressive. It may be worth investigating the hypothesis that “incomplete” CSNB could develop into “complete” CSNB during a lifetime.

Genetic mapping of additional CSNBX families will probably provide information on the chromosomal position of XLRP genes as well. While the gene for XLRP type 3 has been very well localised, the position of the gene for XLRP type 2 has not yet been clearly established. Since XLRP type 2 is very rare, the genetic mapping of additional CSNBX families may be of crucial value in determining...
the genetic position of this disease gene on the proximal short arm of the X chromosome. Finally, in the course of positional cloning studies of XLRP or CSNBX genes, it may be well worth the effort to screen the DNA of both types of patients for subchromosomal deletions. Such aberrations have previously been proven to be of crucial value in the positional cloning process.

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