1 Genomic quantitative genetics to study evolution in the wild

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22 **Abstract**

23 Quantitative genetic theory provides a means of estimating the evolutionary potential of natural
24 populations. However, this approach was previously only feasible in systems where the genetic
25 relatedness between individuals could be inferred from pedigrees or experimental crosses. The
26 genomic revolution opened up the possibility of obtaining the realized proportion of genome shared
27 among individuals in natural populations of virtually any species, which could promise (more)
28 accurate estimates of quantitative genetic parameters in virtually any species. Such a ‘genomic’
29 quantitative genetics approach relies on fewer assumptions, offers a greater methodological
30 flexibility, and is thus expected to greatly enhance our understanding of evolution in natural
31 populations, for example, in the context of adaptation to environmental change, eco-evolutionary
32 dynamics, and biodiversity conservation.

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35 **Understanding trait evolution in natural populations using quantitative genetics**

36 Reliably predicting responses to selection is essential in animal and plant breeding, for studying
37 evolution in natural populations, and understanding population and species response to rapid
38 anthropogenic environmental change [e.g. 1]. Building on the seminal work of RA Fisher [2], who
39 first proposed to decompose the variation in a continuous trait into genetic and environmental
40 components, breeders and evolutionary biologists have long used quantitative genetic (QG) theory
41 to estimate the heritable proportion of trait variation and co-variation with other traits to predict
42 response to natural and artificial selection [3]. Partitioning the variance in a trait has been possible
43 by assuming a simple model of genetic determinism: each trait is affected by many loci of
44 individually (infinitesimally) small effects, known as the infinitesimal model (see Glossary).
45 Despite this strong, and not necessarily realistic assumption underlying QG [4], it has been very
46 efficient and successful in predicting selection responses in animal and plant breeding over the last
47 century [5].

48 The key requirement for any QG analyses is information about the relatedness among individuals.
49 The resemblance between relatives is the basis for separating the heritable proportion of the trait
50 variance (the additive genetic variance) that determines the response to selection and potential for
51 evolution. The generalization of QG theory to include any kind of relatives in a single statistical
52 framework called the ‘animal model’ has become the major tool in animal breeding to predict the
53 ‘genetic merit’ or breeding value of individuals [6]. The animal model also opened up the
54 possibility to study evolution in the wild, because it was no longer necessary to perform crosses to
55 obtain many pairs of individuals of the same relationship, instead, any degree of relatedness was
56 regarded as informative. Nevertheless, the application of QG in wild populations remained a tool of
57 privilege, limited to long-term studies, where pedigrees were possible to establish from individual
58 observations, biasing the approach towards a handful of mammal and bird species [7].

59 With the genomic revolution, QG moved away from the spotlight, and the focus shifted to searching
60 for the genetic determinants of phenotypic variation, i.e. mapping quantitative trait loci (QTL).
61 Thanks to lower genotyping costs, mapping at the finest possible level was enabled by genome wide
62 association studies (GWAS) that aim to find markers, most often single nucleotide polymorphisms
63 (SNPs), associated with phenotypic variation [e.g. 8]. However, GWAS so far did not meet the high
64 expectations: in many cases, a large part of the phenotypic variance remained unexplained by SNPs
65 that reached genome-wide significance (the so-called missing heritability problem); a problem that
66 is now partly resolved by increased sample sizes [8]. Furthermore, even the largest GWAS studies
67 in humans have not yet reached a plateau of the number of 'risk factors' (significant SNPs)
68 associated with trait variation [8]. Not so surprisingly, GWAS conducted in natural plant and animal
69 populations had so far limited power and often failed to identify SNPs associated with traits
70 [reviewed in 9]; yet the lack of any genome-wide significant loci indicates the absence of major
71 effect loci. Some of these studies instead found that the proportion of variance explained by a
72 chromosome correlated with its size [10, 11], as expected if these traits are under the influence of
73 many loci of small effects. All these findings support that the infinitesimal model of classical QG

theory is a reasonable approximation for most quantitative traits, including life-history, behavior, or morphology [12]. The QG approach also has the advantage of not relying on unknown underlying molecular or functional details of the genetic basis of quantitative traits and directly provides estimates of the evolutionary potential of such traits [13]. In this Opinion paper, we promote a so-called ‘genomic QG’, which makes use of genomic data almost exclusively for estimating the relatedness between individuals, thus opening the door to many applications of QG to decipher the architecture of quantitative traits and their response to selection; questions often off-limits in many natural systems. Genomic QG has the potential to improve our ability to mitigate species’ responses to environmental change [e.g. 1], and contribute to the field of conservation genetics, such as identifying ‘pre-adapted’ individuals in programs of assisted migration to ‘help’ species keep up with climate change by estimating genetic merit of translocated individuals [e.g. 14].

From pedigrees to genome sharing

Relatedness is traditionally based on pedigrees at the base of which individuals are assumed to be unrelated (founders) [15]. Given a pedigree, it can be determined whether two alleles at a particular locus that are identical (by state or IBS) are also identical by descent (IBD) because they were inherited from the same ancestor. Due to Mendelian segregation, there is inherent variation in IBD across loci. For example, full-sibs carry, on average, 50% of their alleles IBD, but the exact percentage of genome sharing and hence their realized relatedness is subject to variation (see Fig.1A). The amount of variation around the expectations depends upon the pedigree-relationship class, and is zero only for parent and their offspring and mono-zygotic twins. Pedigree-based relatedness has well known shortcomings, such as its arbitrariness with respect to a founder population and its imprecision regarding IBD allele sharing [e.g. 16]. Nevertheless, it has been standard in the vast majority of past QG studies.

Even though different relatedness estimators that approximate IBD probabilities have been suggested in the microsatellite era, they had too large sampling errors [e.g. 17], thus it was widely

100 accepted that markers should only be used to check, correct, and complete pedigrees rather than
101 replacing them [18]. With the advent of genomic data (Box 1), it has become possible to estimate
102 the realized proportion of the genome that two individuals share IBD. The proportion of genome
103 shared between all pairs of individuals in a population can be summarized with the genomic
104 relationship matrix (GRM) (see Box 1 for methods).

105 The number of markers necessary to accurately estimate the GRM ultimately depends on the
106 effective number of independently segregating chromosome segments that depend on the effective
107 population size, the recombination rate and the genome size [19] and are also influenced by the
108 mating system and population structure and history, which concertedly determine linkage
109 disequilibrium decay. Generally speaking, species with small genomes need fewer markers than
110 species with big genomes, where the difference between expected and realized relatedness can be
111 high [19, 20]. A greater number of markers is also expected to be necessary in outcrossing
112 populations in comparison to selfing, partially selfing or inbred populations because outcrossing
113 populations generally have large effective population sizes and high recombination rates [21, 20]. It
114 is difficult to provide a quantitative guidance for the required number of markers for a specific
115 study. Thus, performing simulations using characteristics of the studied species and populations,
116 and exploring multiple GRM estimation methods (see Box 1) might be the best approach.

117

118 **The ‘dawn’ of genomic quantitative genetics**

119 For humans and species of economic importance, genomic data became available much earlier than
120 for other species, thus the first genomic QG tools appeared in these fields. Many of these methods
121 have not reached evolutionary and global change biology yet. First, we review these tools, then
122 discuss the initial applications in natural populations.

123

124 *Animal and plant breeding*

Animal breeders traditionally estimated breeding values based on phenotypic scores, such as milk yield, of a focal individual and its relatives, using an animal model (Box 2). With high density SNP chips (Table 1) becoming available for many domesticated species, breeders were the first to suggest the use of the GRM in an animal model to predict genomic breeding values (GEBVs) [22] and also developed other approaches for genomic selection (GS) that even bypass the animal model (see Box 2 for details). GS revolutionized animal breeding and the field has experienced a surge of method development [23]. It was also demonstrated that an animal model coupled with a GRM instead of a pedigree leads more accurate estimates of the variance components than pedigree-based equivalents [24, 23]. Further, most pre-genomic breeding applications ignored estimating non-additive genetic effects because they generally require complex breeding designs or pedigree structures, and also because it was believed that their effects are negligible [e.g. 5]. A handful of recent empirical studies from tree breeders demonstrated that, in comparison to classical pedigree based approaches, the use of a GRM in an animal model can yield substantially more accurate separation of the additive and non-additive components of genetic variance [25, 26].

Human genetics

Human populations cannot be manipulated experimentally, so different 'non-invasive' genomic QG methods have been developed in this field. Most notably, it was suggested to exploit variation in realized genome sharing within a pedigree relationship class to estimate the heritability: Visscher *et al.* [27] proposed this idea within full-sib families, and Yang *et al.* [28] among unrelated individuals. Most importantly, these methods led to more accurate estimates of the variance components. Nevertheless, population parameters such as linkage disequilibrium between marker and quantitative trait loci can lead to different performance of this method in different species or populations. To explore this further, Visscher and Goddard [29] investigated the sampling error in the heritability estimates as a function of the sampling scheme, the sample size and the effective population size, and concluded that overall, more individuals are needed to achieve the same

precision with random population samples in comparison to pedigrees, and higher sample size is required when there is little variance in relatedness such with large effective population size or in expanding populations.

The first 'wild' applications

Applications of the genomic QG approach in wild populations have been lagging behind breeding and human genetics for obvious genomic resource reasons, even though the idea to use molecular markers to estimate heritability in natural populations had been proposed back in 1996 by Ritland [31]. Most early applications, based on a few microsatellites, yielded estimates too imprecise and often out of range [30, 31]. Recently, with dense marker panels becoming accessible in some wild pedigreed populations, it has been shown that marker-based relatedness estimates agreed with those from pedigrees (see Fig.1A and B) and also that heritability estimates based on high-density markers and pedigrees agree (Soay sheep (*Ovis aries*) [32], great tits (*Parus major*) [10]). Some empirical studies also investigated how many markers are required to estimate the heritability by comparing estimates from a decreasing proportion of the available loci. For example, in the selfing plant species, *Medicago truncatula*, heritability estimates with 25k SNPs (i.e. one marker every 10 Kb, which approximates the distance of linkage disequilibrium decay) were nearly unbiased in comparison to those obtained with >5 million SNPs (Fig.1C). Similarly, for a small island population of a wild ungulate, Soay sheep, heritability with 10k SNPs yielded unbiased estimates of the heritability obtained with 33k SNPs across several traits (Fig.1D).

The genomic QG toolbox for ecology and evolution

Bringing QG to the wild by using the pedigree-based animal model has greatly advanced our understanding of evolution [7]. Today, the acquisition of high-density marker data is possible in virtually any species (Table 1), which opens up the possibility to apply the QG research program in

176 the absence of a pedigree or controlled cross in any population of any species. Genomic QG also
177 offers a greater methodological flexibility than classical QG and many evolutionary and global
178 change research questions could benefit from it.

179

180 *Genomic estimated breeding values*

181 GEBVs could have many potential uses outside animal and plant breeding, such as in the context of
182 conservation biology to identify suitable individuals for assisted migration or release programs. The
183 objective of assisted migration is to ‘help’ species migrating to keep up with the pace of climate
184 change, and it is becoming common practice for forest trees [14]. It has been suggested that GEBVs
185 can be combined with gene-environment association analysis to predict phenotypes and identify
186 changes in performance along environmental gradients [33]; Box 3 provides a proof-of-concept
187 example of this approach in *Arabidopsis thaliana*. Since GS is based on a statistical model built on
188 data from a ‘training population’ (Box 2); in order to obtain reliable predictions of the breeding
189 values; it seems necessary to include individuals across the full environmental range over that
190 predictions are desired. A further potential use of GEBVs is in breeding programs of endangered
191 species, where genomic data could help to optimize breeding of remaining individuals to minimize
192 inbreeding or outbreeding depression [34]. Releasing individuals bred in captivity into the wild is
193 often used to restore extinct populations or help threatened populations to survive, but programs
194 might fail if individuals are maladapted to the new environment [e.g. 35]. Identifying potentially
195 well-adapted individuals based on their GEBVs, as described in Box 2, could be an option, or
196 combined with allele specific genetic rescue, e.g. when disease resistance genes are known [see
197 examples in 36]. As pointed out above, it remains unclear whether estimating GEBVs will be
198 generally feasible in natural populations, but in an outbreeding tree species GEBVs estimated with
199 reasonable accuracies [37]. The fact that QG estimates based on genomic markers agreed with
200 ‘standard’ QG estimates in the studies on mammals, birds and plants [10, 38, 32] also indicates that
201 it should be possible.

202

203 *Realized genome sharing to quantify fitness*

204 The GRM is just the first step in genomic QG, but it can also be used in another innovative way in
205 evolutionary biology: The average relatedness of an individual with the population, estimated from
206 a GRM, could be used as a measure of inclusive fitness integrated across the current population as
207 individuals whose ancestors produced many offspring should have a higher than average relatedness
208 with the population. Such a fitness measure could also be interesting for behavioral ecology, where
209 estimating relatedness among interacting individuals is necessary to understand social behaviors.
210 Here, the average genomic relatedness of an individual could better estimate its inclusive fitness.
211 Particularly, relatedness estimates based on coalescence could be adapted here [see also 39 and Box
212 1]. Estimating reproductive success is a challenging endeavor in wild populations for several
213 reasons [40] but could potentially be tackled by estimating GRMs.

214

215 *Evolutionary potential in the wild*

216 Evolutionary potential is a scaled measure of the additive genetic variance (V_A); it is the heritability
217 when it is scaled with the total phenotypic variance and evolvability when it is scaled by the trait
218 mean [41]. Bringing genomic QG to the wild will allow assessing the evolutionary potential of a
219 population *in situ*, which is a considerable advancement because it estimates the additive genetic
220 variance and covariance of phenotypic traits under the conditions in which natural selection is
221 acting, which is not the case when using common garden experiments. Such common garden
222 estimates are not guaranteed to apply under natural conditions as V_A might vary across
223 environments and strongly depends on population size and inbreeding levels [reviewed in 42].
224 Estimates of trait heritability in natural populations, however, are also prone to biases due to plastic
225 responses to environmental variables [see 43 for a recent review]. Consequently, a good
226 understanding of environmental but also individual-level variables, such as age or body condition, that

affect the trait is necessary to account for this ‘environmental’ variation by including the relevant variables in the model. Alternatively, if the exact variables responsible for plastic trait variation are unknown or difficult to estimate but are known to vary spatially or temporally, appropriately modelling this spatial or temporal variation is a suitable approach. For example, in the case of an environmental variable that varies in discrete ‘units’, such as the effect of spring climate on timing of flowering, breeding or migration, fitting the temporal or spatial ‘unit’ as a (fixed or random) factor would be appropriate. If variation is more gradual, fitting an appropriate autocorrelation in the model can remove this plastic variation in traits [e.g. 44]. Another approach to remove confounding effects between genetic and environmental similarities is to estimate QG parameters from variation in relatedness within families, such as full-sibs [27], which could be possible in clutches of birds, fish or insects or half-sibs in plants [e.g. 45]. These *in situ* estimates of the evolutionary potential can be useful in many fields of ecology, evolution and conservation biology. Climate change induced new selective pressures such as increased drought, heat or seasonal shifts, which have been reported in many ecosystems but for most species we lack *in situ* or even any estimates of V_A at ecologically key traits including phenology or drought tolerance [e.g. 46, 47]. Climate change is also expected to drive changes in community assemblages due to different dispersal abilities at different species, as has been shown for alpine plants [48]. *In situ* evaluation of the evolutionary potential of key life-history traits is necessary to assess whether these species will be able to adapt to the changed competition regime.

Estimates of V_A could also be particularly useful for models of eco-evolutionary dynamics (Hendry 2016). For example, Ellner [49] suggested a framework to partition change in a population into contributions of evolution, non-heritable trait change and environmental. To date, this theoretical framework has been limited to experimental systems, but with genomic QG it could be applied in natural settings.

With the decreasing costs of genotyping, measuring phenotypes of individuals can become the limiting factor for QG studies in natural populations. Although the variance in (genomic) breeding

values is not V_A [see e.g. 50], it could be used as a proxy for it, which would allow prediction of evolutionary potential in new populations without the need to phenotype individuals. However, genotype-by-environment interactions might limit the application of this approach to species where reaction norms are known across environmental gradients.

Predicting response to selection

The evolutionary potential informs about the capacity of a population to respond to selection. Knowledge of V_A for a trait or for fitness directly applies to predictive QG models of eco-evolutionary dynamics. This is especially relevant in the context of predicting shifts of species' ranges caused by climate change, and many voices have called for the development of eco-evolutionary forecasting models in that context [e.g. 51]. Quantitative genetics theory provides a well formed framework for predicting eco-evolutionary dynamics in single or among competitively interacting species [e.g. 52]. Current research efforts provide practical examples of implementation of this framework, together with modeling of species ecological niche distributions, to predict changes of species' distribution in *Drosophilid* flies in Australia [53] or endemic alpine plants in Austria [54]. However, this approach is data hungry and needs estimates of V_A for climate-adaptation traits and the selection gradients acting on them. There is hope that by using genomic QG tools such as environment-related GEBVs in natural populations (Box 3) we will be able to alleviate the dearth of estimates of QG parameters necessary to correctly forecast the fate of natural species facing a warming climate.

Conclusions

Genomic QG has the potential to advance our understanding of evolutionary processes in natural populations in several ways. Its main advantage is that we no longer need pedigrees or breeding experiments for QG analysis, which means that we can study additive genetic (co)variances, but

278 also selection, in virtually any species in a natural setting. Genomic QG parameters estimates can be
279 more precise than pedigree-based estimates, because they have fewer assumptions and offer a
280 greater methodological flexibility. This ability to conduct quantitative genetic analyses in natural
281 populations could allow more accurate predictions of species' persistence under climate change and
282 enable informed conservation strategies. As we pointed out, the feasibility of this approach will
283 partly depend on the specific study system. The relative ease with which high-density markers can
284 be obtained make us cautiously optimistic about the potential of marker-based quantitative genetics
285 to considerably contribute to our understanding of evolutionary processes in natural populations and
286 mitigating climate change.

287

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448 Table 1 Overview over high-throughput genotyping approaches and sequencing approaches that are potentially suitable to estimate relatedness.

Data type	Genomic resources needed	Pros	Cons	Cost	Examples of studies applying this approach to calculate the GRM
SNP chip	Reference genome	“Ready to use” - No bioinformatics work	Availability mostly limited to model and breeding systems, and possibly close relatives. Ascertainment bias: new populations can have different polymorphisms [55]	Low, if chip available	animal and plant breeding [24, 23], human genetics [28], wild species [32]
RRL- or RAD-sequencing	None, but improved if there is a reference genome	Flexibility of experimental design assisted by software predictions on number of markers. Established bioinformatics for data	Bias on relatedness estimates due to allele dropout and high proportion of missing data [56]	Medium	[56, 57]

		analysis with easy-to-use pipelines			
Whole-genome sequencing	Reference genome	High-confidence genome-wide information and use of established bioinformatics tools	High costs for large genomes, and can yield excessive amount of information [38]	Very high	[58, 38]
Low-coverage whole-genome sequencing	Reference genome	Dense genome representation including causal variants that can be suitable for parallel association studies.	Requires bioinformatics knowledge to use software building on probabilistic framework.	High	[59, 60]
Sequence or Exome capture	Reference genome of closely related species or	Targeted sequencing of selected loci, which can include causal variants	Requires high-quality genomic resources for bait design and to minimize allele bias [61]	High	[62]

	transcriptome	suitable for parallel association studies.			
Genome skimming	Reference genome	Minimal sequencing effort to survey high-copy regions of the genome, typically organellar and ribosomal DNA	No studies yet built a GRM. Complex evolution of high-copy nuclear loci can limit applicability in natural populations.	Low	

BOXES

BOX 1 Methods to estimate the genomic relationship matrix (GRM)

1. Approximate identity-by-descent (IBD) probabilities at single locus. This approach includes a whole class of maximum-likelihood [e.g. 63, 60] and method of moments estimators [see 17 for an overview].

2. Cumulative excess of recent coalescence. A coalescent tree is used to trace the origin of alleles in the current population back to a most recent common ancestor (MRCA). Rousset [39], suggested to define kinship coefficients as the degree of excess coalescent time to their MRCA at each independently segregating segments of the genome, however, no estimator or software have been proposed yet.

3. Average allelic correlation (a.k.a. unified additive relationship). This is the most standard class of approaches to estimate the GRM in animal and plant breeding and human genetics (see Table I for software). In breeding, the method appeared with the uprise of genomic selection (GS), and calculated as the correlation between genotypes of two individuals across all loci [64]. In human genetics, the approach appeared in Yang et al. [28], and was explained in Powell et al. [65]. The approach circumvents the problem of the arbitrarily reference population by defining it as the current population. As a result, IBD is no longer a probability, but a correlation coefficient that can be used to predict if a gamete carries the same allele as another irrespective of its origin (i.e. IBS). Adjustment for linkage disequilibrium (LD) been proposed by [16], which can lead to improved estimates, especially with sparse SNP data [66]. When using dense SNP data, LD-adjustment attributes too much weight to the SNPs with low minor allele frequencies.

4. Haplotype sharing. Haplotype sharing approaches require both marker data and a genetic map [e.g. 67]. A common approach in GS is to use dense SNP chip on a few key individuals to identify haplotypes, and a sparse chip on the rest of the individuals [see review by 23].

474 Table I. Examples of software to estimate the GRM

Soft ware	PLINK (v1.07)	GCTA	LDAC	BEAGLE
Meth od	1	3	3	4
Refer ence (soft ware)	[68]	[69]	[16]	[67]
Webs ite	http://zzz.bwh.harvard.edu/plink/	http://cnsgenomics.com/software/gcta/	http://dougspeed.com/ldac/	http://faculty.washington.edu/browning/beagle/beagle.html

475

476

477 **BOX 2 Two key statistical models to estimate QG parameters using genomic data**

478 The two main types of statistical models dominate the QG literature and both can make use of
 479 genomic data. In the animal model, the pedigree can be replaced with the GRM, and in genome
 480 regressions, genomic data is a necessary ingredient.

481

	Animal model	Genome regression
Key reference	Henderson (1975)	Meuwissen, Hayes, Goddard (2001)
The model	$y_i = \mu + a_i + e_i$ where a_i is the breeding value of individual i. Since a_i 's are unknown, they are estimated from the covariance between relatives in additive genetic effects given by $2 \times \text{kinship} \times V_A$, thus using a pedigree or a GRM	$y_i = \mu + \sum (SNP_{i,j} * g_j) + e_i$ where $SNP_{i,j}$ is the genotype of individual i at loci j, and g_j is the effect of the SNP on the phenotype
explanation of common model terms	y_i is a trait value of individual i, μ is the population mean, and e_i is the residual term	
Breeding value (BV) estimation	Using the best linear unbiased predictor (BLUP) in a mixed effects model	BLUPs assuming equal variances across SNPs is possible, but Bayesian methods generally work better

482

483 **About the animal model**

484 Animal model is most often used to estimate variance components, such as the additive genetic
485 variance, which serves to estimate the heritability of traits or genetic correlations between traits.
486 Breeding values are the estimated random effects and can be used to characterize individuals,
487 however, several authors warn not to use them in further statistical analysis without incorporating
488 their estimated standard errors [e.g. 50], e.g. in a hierarchical Bayesian framework. Generally large
489 pedigrees are required but it can also be used with a common garden experimental data.

490

491 **About genome regressions**

492 Genome regressions are principally used in a prediction context to help breeders identify superior
493 individuals. Thus, the model is fitted in a training population to estimate breeding values of
494 individuals of unknown trait value in a candidate population (such as phenotyping milk yield at an
495 early age or at a male cow). GEBVs are generally much more accurate than EBVs. Particularly high
496 accuracy is expected with a large number of individuals in the training population, for a highly
497 heritable trait, and when the number of segregating chromosome segments is small (i.e. small
498 genome and small effective population size). GS does not work well across breeds. Instead, the best
499 performance is achieved when individuals in the candidate population are related to individuals in
500 the training population.

501

Box 3 Using genomic relatedness and home environments to identify locally adapted genotypes

A common challenge in conservation genetics is to identify locally-adapted genotypes for a given site. Here, we demonstrate that GRMs for wild accessions collected across diverse climates (‘ecotypes’) can be applied to predict which genotypes are adapted to cold sites and which are adapted to warm sites. Our approach relies on a three-way correlation between genomic similarity (GRMs), similarity in home environments (winter cold), and similarity in locally adaptive traits (unobserved and predicted). As such, correlation between population structure and environmental gradients causing local adaptation is not a problem for our approach, rather this correlation is leveraged to predict adaptive trait variation. By contrast, population structure-environment correlation is problematic for genome-environment association studies that seek to identify individual causal loci involved in local adaptation. We reanalyzed published data on *Arabidopsis thaliana* [70-72] and global climate [73]. We used >200k SNPs [72] to estimate a GRM based on average allelic correlation (Box 1). We fitted an animal model where an ecotype’s native winter cold environment (minimum winter temperature) was used as ‘phenotype’ and modeled as a function of the GRM with 10-fold cross validation to predict native temperatures as GEBVs [74]. Based on the assumption that populations are locally adapted along temperature gradients, these GEBVs can also be interpreted as predictions of a ‘latent cold-adapted phenotype’, *i.e.* an unobserved phenotype assumed to confer high fitness in particular temperatures. We next show that these predictions closely correspond to which genotypes are locally adapted along temperature gradients. Although here we validated our predictions with experimental data, note that no experiments were used to generate our predictions; predictions are based solely on the relationship between GRM and native environments. Individual relative fitness of 157 ecotypes was measured in four common gardens by Fournier-Level et al. [71]. In each garden, we regressed relative fitness against predicted latent cold-adapted phenotypes to identify the expected most fit latent phenotype at each site (Fig. I, data from two sites shown in insets, curve shows fitted quadratic regression).

528 The fittest latent cold-adapted phenotype at each site varied closely with differences in winter cold
529 among sites (central panel, $R^2 = 0.92$). This relationship indicates that the GRM association with
530 native environments can be used to predict which genotypes are adapted to cold sites and which are
531 adapted to warm sites, information that might guide conservation and restoration actions.

532

Glossary

Infinitesimal model: The infinitesimal model assumes that the genetic component of a trait is determined by an infinite number of unlinked genes that each has an infinitesimally small, additive effect. This will lead to continuous genetic variation in the trait and this model is hence generally used to model so-called ‘quantitative’ or ‘complex’ traits. It is, however, also applicable to discrete traits, as e.g. litter size, and even binary traits if trait expression depends on one or more thresholds that the summed allelic effects need to pass.

Heritability: The narrow-sense heritability (h^2) is the ratio of the additive genetic variance (V_A) to the total phenotypic variance (V_P) of a trait: $h^2 = \frac{V_A}{V_P}$. The narrow-sense heritability is used to predict the response to selection of a trait when using the breeders’ equation. When only the whole genetic variance (V_G) is known, which also includes non-additive genetic effects (dominance and epistasis), broad-sense heritability (H^2) is estimated.

Evolvability can be defined in the broader and narrower sense. In the broader sense it simply is the ability of a trait to evolve (including from novel mutations). In the narrower sense it is the additive genetic variance scaled by the trait mean: $volvability = \frac{V_A}{\text{trait mean}^2}$, or the additive genetic coefficient of variation (CV_A): $CV_A = \frac{V_A}{\text{trait mean}}$.

Quantitative trait locus: A quantitative trait locus (QTL) is the genomic section, i.e. a locus, that contribute to the variation (together with other QTL) of a quantitative trait, i.e. a trait that shows continuous phenotypic variation.

Genome-wide association study: A genome-wide association study (GWAS) aims to identify loci determining a trait by testing for associations between molecular markers and trait values. In the common case when SNP chips are used, the underlying assumption is that the markers are in tight linkage with or within the loci responsible for trait variation.

557 **Single nucleotide polymorphism:** A single nucleotide polymorphism (SNP) is a variation at a
558 specific genomic position. For example, at a specific position in a genome, the base C appears in
559 most individuals, but in a minority the base A appears. This specific SNP would have the two
560 nucleotide variations – C or A – as its alleles.

561 **Identity-by-descent:** A genomic section is identical by descent (IBD) in two individuals if they
562 have inherited it from a common ancestor.

563 **Identity-by-state:** A genomic section is identical by state (IBS) in two individuals if it has the same
564 nucleotide sequence in these two individuals. IBS can arise by chance (mutation) or because the
565 individuals inherited it from the same ancestor, in which case it would also be identical by descent.

566 **Realized relatedness:** The relatedness of two individuals calculated from a pedigree (kinship) is
567 their expected genetic similarity (or proportion of genome sharing). There is, however, variation
568 around this expectation (except for parent-offspring pairs and monozygotic twins) because of the
569 segregation of independent genome segments during meiosis (mendelian noise). This variation is
570 lowered by more independently segregating segments[e.g. 27, 19]. An interesting consequence of
571 this is that for species with ‘small’ genomes, i.e. with fewer independently segregating segments,
572 QG estimates based on realized relatedness will be more precise than estimates based on expected
573 relatedness.

574 **Genomic relationship matrix:** The pedigree-based relatedness matrix, often denoted A , contains
575 all pairwise relatedness estimates (which are two time the kinship coefficient θ) in the off-diagonals
576 and the individuals inbreeding coefficient in the diagonal. The genomic relatedness matrix (GRM),
577 often also denoted G , is the same but with the relatedness estimates derived from genomic markers.
578 Various approaches to estimate relatedness from markers exists (see Box 1) and GRMs can be
579 constructed with any of them.

580 **Linkage disequilibrium** is the non-random association of alleles at different loci. Loci are in
581 linkage disequilibrium, or linked, if certain combinations of alleles occur more often than expected

582 by chance. Linkage disequilibrium can be caused by a multitude of factors, including selection,
583 recombination rate, mating system, population structure or physical proximity in the genome.

584 **Non-additive effects** include dominance and epistasis. Dominance means that the effect of one
585 allele masks (is ‘dominant’ over) the effect of the other (recessive) allele. Incomplete and co-
586 dominance are cases when the dominant allele is not completely masking the recessive allele and
587 the heterozygous phenotype is somewhat intermediate between the homozygous dominant and
588 homozygous recessive phenotype. Epistasis, or gene-gene interactions, occurs when the allelic
589 effect at one locus depend on the alleles of one or more other loci, the genetic background.

590 **Assisted migration:** Many species have limited dispersal abilities or their dispersal is impaired by
591 habitat fragmentation, which means that they cannot reach other, isolated populations or currently
592 unoccupied but suitable habitat. In assisted migration, or assisted gene flow, individuals are
593 artificially moved from one population to improve another population’s probability to persist or to
594 colonize novel but suitable habitat, e.g., in the context of climate change.

595

Figure caption

Fig.1 Genomic relatedness and heritability estimates. (A) Relationship between marker-based and pedigree relatedness in one of the wild study populations with the deepest pedigree, the St. Kilda soay sheep population. Realized or ‘genomic’ relatedness, estimated from ca. 37000 SNP markers, is plotted against IBD relatedness, estimated from the pedigree. The variation of realized relatedness estimates with the same IBD relatedness class, seen as vertical spread, is due to Mendelian segregation (and sampling error). (B) shows, for the same population, that the correlation between pedigree and marker-based relatedness estimates increases with the number of SNPs used to estimate the GRM. (C) and (D) show the effect of number of markers on heritability estimates in *Medicago truncatula* and soay sheep, respectively. Boxes and whiskers indicate the variation in estimates from the repeated samples of the reduced data sets. When few markers are used to estimate the GRM heritability, estimates are down-ward biased. Increasing the number of markers reduces this bias, but, at some point, adding more markers has little effect on the estimate anymore, only on its variation. In (B) and (D) the x-axis shows the proportion of SNPs used in relation to the complete data set and the absolute numbers are 926, 1852, 3704, 11111, 18518, 25926, respectively. (A), (B) and (D) re-drawn after [32]; (C) re-drawn after [38].